

Ethyl 2-aminoxazole-5-carboxylate

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Key indicators

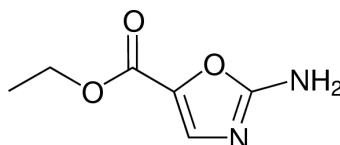
Single-crystal X-ray study
 $T = 150\text{ K}$
Mean $\sigma(\text{C}-\text{C}) = 0.003\text{ \AA}$
 R factor = 0.050
 wR factor = 0.126
Data-to-parameter ratio = 11.9

For details of how these key indicators were automatically derived from the article, see <http://journals.iucr.org/e>.

The crystal structure of ethyl 2-aminoxazole-5-carboxylate, $\text{C}_6\text{H}_8\text{N}_2\text{O}_3$, consists of planar sheets internally connected by intermolecular hydrogen bonding running parallel to the (120) plane. Interactions between the sheets are limited to dipole–dipole interactions between antiparallel carbonyl groups.

Comment

Ethyl 2-aminoxazole-5-carboxylate, (I), was prepared as part of an ongoing investigation of anticancer antibiotic drugs such as distamycin and netropsin (lexitropsins; see Khalaf *et al.*, 2000). Since these drugs exhibit biological activity by binding to specific (A/T)₄ sequences in the minor groove of DNA, several analogues with subtly differing binding sites were prepared and tested for activity (Fishleigh *et al.*, 2000). The antiviral activity of (I) was tested *in vitro* by inhibition of plaque formation in cell cultures infected with several RNA and DNA viruses, including Newcastle–Disease and Pseudorabies virus (Ulbricht, 1987).



(I)

The molecular structure of (I) is essentially planar with only the atoms of the ethyl tail showing any significant deviation from the plane defined by the atoms of the five-membered ring [C5 0.163 (6) Å and C6 0.326 (8) Å]. Sheets of coplanar molecules lie parallel to the (120) plane. These sheets are held together internally by in-plane intermolecular hydrogen bonding involving the amine H atoms and all other available acceptor atoms [N2–H2...N1ⁱ 2.09 (3), N2–H1...O2ⁱⁱ 2.14 (3) and N2–H1...O1ⁱⁱⁱ 2.63 (3) Å for the H...A distances; symmetry codes: (i) $-x, -y, -z + 2$; (ii) $-x, -y, -z + 1$]. There are dipole–dipole interactions along the *b* direction and between these sheets involving the carbonyl group [O2...C4ⁱⁱⁱ 3.545 (3) Å; symmetry code: (iii) $-x + 1, -y, -z + 1$], but no short π interactions involving the ring atoms.

A search of the Cambridge Structural Database (Allen & Kennard, 1993) found no primary amine analogues to (I) but did find a series of secondary and tertiary amines which had been synthesized as aldose reductase inhibitors (Shibata *et al.*, 1982; Ishida *et al.*, 1991). (I) shows several differences in intraring bond distances as compared to these compounds. Most notable is the considerable lengthening of the C1–N1 bond

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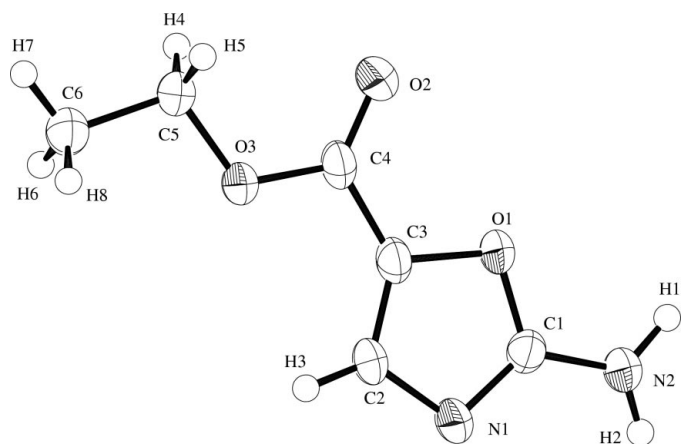


Figure 1
ORTEPII (Johnson, 1976) view of (I) with non-H atoms as 50% ellipsoids and H atoms as small spheres of arbitrary size.

[1.316 (3) Å *cf.* a range of 1.263–1.294 Å for the five related fragments found] and a shortening of the C2–N1 bond [1.378 (3) Å *cf.* 1.385–1.410 Å]. The bond lengths appear to show that the N2 amine lone pair is considerably conjugated through the ring.

Experimental

Sodium ethoxide (2.44 g, 0.33 mol) was added to diethyl ether (500 ml) and cooled to 273 K with stirring. Ethyl chloroacetate (40.5 g, 0.33 mole) and ethyl formate (24.3 g, 0.33 mol) were added to the reaction mixture over 2 h. The stirring continued overnight at room temperature. The reaction mixture was diluted with ice/water (500 ml) until the sodium salt dissolved. The diethyl ether layer was separated and the water layer was acidified with concentrated HCl and cooled. The precipitated ester was filtered off. After work-up, 43.70 g of crude product was distilled at 5 mmHg (333–338 K). The product, ethyl 2-chloro-3-oxopropanoate C₅H₇ClO₃, recrystallized on standing (22.44 g, 45%) as white needles (m.p. 288–293 K). To the above starting material, urea (8.95 g, 0.149 mol) was added followed by 100 ml of ethanol. This mixture was refluxed for 2 h. Ethanol was removed under reduced pressure and then a sodium bicarbonate solution (10%) was added until effervescence ceased. Diethyl ether was added and 20.98 g of ethyl (2*Z*)-3-[(aminocarbonyl)amino]-2-chloro-2-propenoate, C₆H₉ClN₂O₃, was removed by filtration (m.p. 484–487 K). The diethyl ether layer was dried over Na₂SO₄ and then the solvent was removed under reduced pressure to leave a green oil. Purification by flash silica-gel column chromatography using ethyl acetate/*n*-hexane (1:2, *R_F* = 0.2) afforded crystalline (I) (1.72 g, 7.4% yield), m.p. 425–428 K (literature m.p. 428 K; Dornow *et al.*, 1953). ¹H NMR (CDCl₃): 1.35–1.36 (3H, *t*, CH₃); 4.31–4.36 (2H, *q*, CH₂); 5.33 (2H, *s*, NH₂); 7.48 (1H, oxazole); IR (KBr): 3387, 3119, 1715, 1676, 1568, 1408, 1376, 1325, 1261, 1178, 1146 cm⁻¹.

Crystal data

C₆H₈N₂O₃
M_r = 156.14
 Triclinic, *P* $\bar{1}$
a = 6.1295 (6) Å
b = 7.8227 (6) Å
c = 8.3961 (8) Å
 α = 113.592 (7)°
 β = 93.831 (4)°
 γ = 95.557 (4)°
V = 364.74 (6) Å³

Z = 2
D_x = 1.422 Mg m⁻³
 Mo *K*α radiation
 Cell parameters from 1086 reflections
 θ = 1.0–26.0°
 μ = 0.12 mm⁻¹
T = 150 (2) K
 Cut needle, colourless
 0.20 × 0.15 × 0.08 mm

Data collection

Nonius KappaCCD diffractometer
 φ and ω scans to fill Ewald sphere
 3501 measured reflections
 1300 independent reflections
 905 reflections with *I* > 2σ(*I*)

R_{int} = 0.064
 θ_{\max} = 25.3°
h = -7 → 7
k = -9 → 9
l = -10 → 9

Refinement

Refinement on *F*²
R [*F*² > 2σ(*F*²)] = 0.050
wR (*F*²) = 0.126
S = 1.06
 1300 reflections
 109 parameters
 H atoms treated by a mixture of independent and constrained refinement

$w = 1/[\sigma^2(F_o^2) + (0.0599P)^2 + 0.0404P]$
 where $P = (F_o^2 + 2F_c^2)/3$
 $(\Delta/\sigma)_{\max} = 0.001$
 $\Delta\rho_{\max} = 0.21 \text{ e \AA}^{-3}$
 $\Delta\rho_{\min} = -0.20 \text{ e \AA}^{-3}$

The H1 and H2 atoms of the amine group were placed as found in difference syntheses and refined isotropically. All other H atoms were placed in calculated positions and refined in riding modes.

Data collection: *DENZO* (Otwinowski & Minor, 1997) and *COLLECT* (Hooft, 1988); cell refinement: *DENZO* and *COLLECT*; data reduction: *DENZO* and *COLLECT*; program(s) used to solve structure: *SHELXS97* (Sheldrick, 1997); program(s) used to refine structure: *SHELXL97* (Sheldrick, 1997); molecular graphics: *ORTEPII* (Johnson, 1976); software used to prepare material for publication: *SHELXL97*.

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